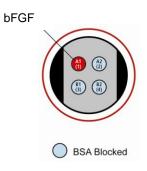
MSD® 96-Well MULTI-ARRAY® Human bFGF Assay

The following assay protocol has been optimized for analysis of basic fibroblast growth factor (bFGF) in human serum and plasma samples.

		Storage
MSD	Materials	
	Read Buffer T (4X)	RT
	Blocker C	2-8 °C
	MULTI-SPOT® 96-well 4 Spot Human bFGF Plate	2-8 °C
	SULFO-TAG™ Anti-hbFGF Antibody (100X)¹	2-8 °C
	Diluent 7	≤-10 °C
	Diluent 8	≤-10 °C
	Diluent 9	≤-10 °C
	Human bFGF Calibrator (1 µg/mL)	≤-70 °C



The ${\sf SECTOR}^{\it @}$ Imager data file will identify spots according to their well location, not by the coated capture antibody name.

FOR RESEARCH USE ONLY.
NOT FOR USE IN DIAGNOSTIC OR THERAPEUTIC PROCEDURES.



¹ Some SULFO-TAG labeled detection antibodies may be light-sensitive, so they should be stored in the dark.

Notes:

Other Materials & Equipment (not supplied)

- Deionized water for diluting Read Buffer
- □ Phosphate buffered saline + 0.05% Tween-20(PBS) for plate washing
- □ Adhesive plate seals
- □ Microtiter plate shaker
- □ Plate washer or other efficient multi-channel pipetting equipment for washing 96-well plates
- Appropriate liquid handling equipment for desired throughput that must accurately dispense 25 and 150 μL into a 96-well microplate

Protocol at a Glance

The protocol can be completed in approximately 5.5 hours if each reagent is prepared during the preceding incubation. This time can reduced to 4.5 hours if the blocking reagent is added the night before.

- **Step 1.** Add Blocking Solution, incubate 1-2 hour, wash. (alternatively, block plates overnight at 4 °C).
- Step 2. Add 25 μL of Diluent 7. Add 25 μL of Samples or Calibrator, incubate 2 hours, wash.
- **Step 3.** Add 25 μL of Detection Antibody, incubate 2 hours, wash.
- **Step 4.** Add 150 µL of Read Buffer, read plate and analyze data.

Read the entire detailed instructions before beginning work.

The assay protocol was optimized for serum and plasma samples. For significantly different sample matrices, it is recommended to use a calibrator diluent that is similar to the sample matrix (e.g. lysis buffer + carrier protein).

Preparation Instructions

Thaw Diluents

Thaw Diluent 7, Diluent 8, and Diluent 9. Vortex briefly. If there is a precipitate, mix gently and warm to room temperature to dissolve. Diluents are stable at 4 °C for one week.

Prepare Calibrator dilutions:

1. Determine how many Calibrator levels and replicates will be tested in the experiment. Each well will require 25 μL of Calibrator. Thaw Diluent 9 and one vial of Human bFGF Calibrator (1μg/mL). Vortex briefly. Prepare the required Calibrator dilution series using Diluent 9.



Notes:	N	O	te	s:
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- 2. A recommended Calibrator dilution procedure is listed below for 4 replicates of 7 Calibrator concentrations spanning a wide range, plus 1 zero-Calibrator point.
 - Prepare 1000 μL of bFGF Calibrator at a concentration of 10000 pg/mL by adding 10 μL of the bFGF Calibrator at 1 μg/mL to 990 μL Diluent 9.
 - ii. Prepare 6 additional 1:4 serial dilutions, beginning with the bFGF Calibrator at 10000 pg/mL, by adding 50 μL of the Calibrator to 150 μL Diluent 9.
 - iii. This will create 7 Calibrators with 10000, 2500, 625, 156, 39, 9.8 and 2.4 pg/mL. The recommended 8th dilution is Diluent 9 alone (e.g. zero Calibrator).
- 3. Calibrators are stable at room temperature for a few hours. The Calibrator stock solution is stable for one day at 4 °C and for one additional freeze-thaw cycle. Diluent 9 is stable for one week at 4 °C.

Prepare the 1X Detection Antibody Solution

- a) In a 15 mL tube combine:
 - \square 30 μ L of 100X SULFO-TAG Anti-hbFGF-1 Antibody
 - □ 2.97 mL of Diluent 8
- b) This will yield 3 mL of diluted Detection Antibody Solution at the working concentration with sufficient volume for one plate.

Detection Antibody Solution is stable at RT for a few hours.

Dilute Read Buffer:

In a 50 mL tube combine (per plate):

- □ 5 mL 4X Read Buffer T
- □ 15 mL deionized water



Assay Protocol

Notes:

Begin with a MULTI-ARRAY 96-well 4 Spot Human bFGF plate. No pre-treatment is necessary.

1. Add 150 μ L/well of Blocker C and incubate at room temperature for 1-2 hours or overnight at 4 °C.

Plates may also be blocked overnight at 4°C and stored for up to a week with Blocker C.

- 2. Wash plates 3 times with phosphate buffered saline + 0.05% Tween-20 (PBS-T).
- 3. Add 25 μ L/well of Diluent 7.
- 4. Add 25 μ L/well Calibrator or sample and incubate at room temperature with shaking for 2 hours.
- 5. Wash plates 3 times with PBS-T.
- 6. Add 25 μ L/well of 1X Detection Antibody Solution and incubate at room temperature with shaking for 2 hours.
- 7. Wash plates 3 times with PBS-T.
- 8. Prepare SECTOR Imager so that plate can be read immediately after Read Buffer addition.
- 9. Add 150 µL/well 1X Read Buffer T.
- 10. Analyze immediately with SECTOR Imager.

Avoid bubbles while adding the Read Buffer; it will interfere with accurate reading of the plate.

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