

MESO SCALE DISCOVERY

MULTI-SPOT Assay System

Whole Cell Lysate Kit - Phospho (Ser 112)/ Total BAD Assay

| | |
|-----------|-----------|
| K15103D-1 | K11103D-1 |
| K15103D-2 | K11103D-2 |
| K15103D-3 | K11103D-3 |



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MSD MULTI-SPOT Biomarker Detection

Whole Cell Lysate Kit

Phospho (Ser 112)/ Total BAD

This package insert must be read in its entirety before using this product.

V8 2010Jul

**FOR RESEARCH USE ONLY.
NOT FOR USE IN DIAGNOSTIC PROCEDURES.**

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Ordering Information

ordering information

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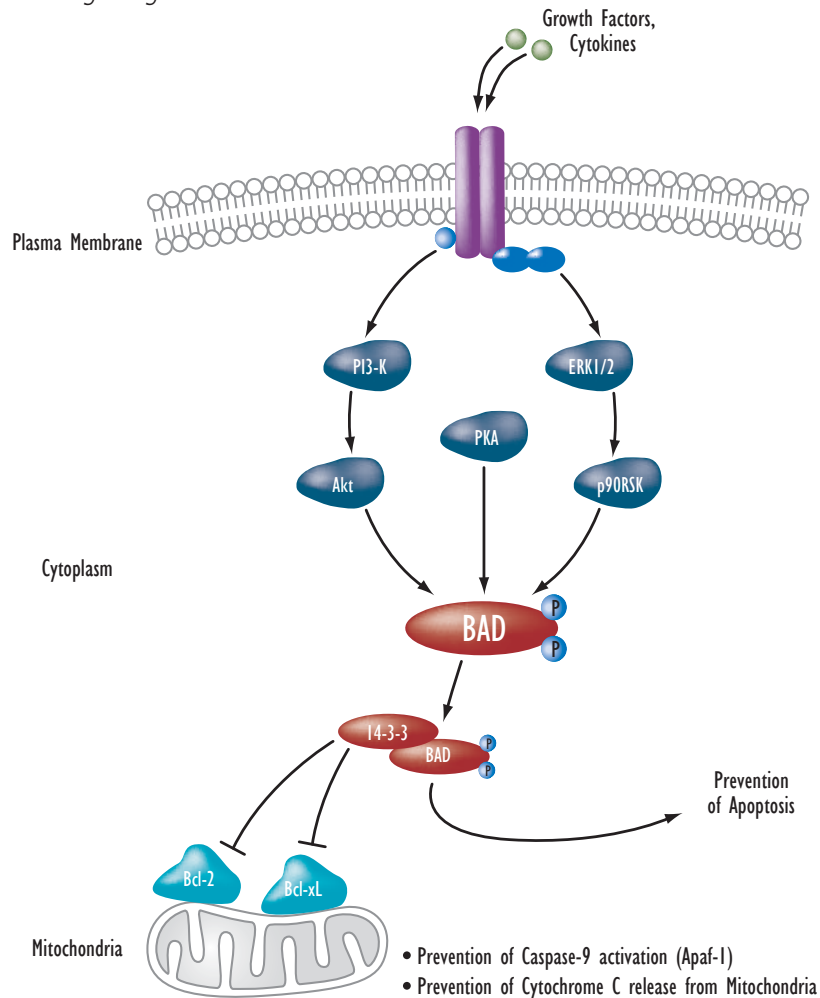
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Introduction

introduction

A member of the pro-apoptotic Bcl-2 family of proteins, BAD functions by displacing the binding of Bax to Bcl-2 and Bcl-xL, causing cell death by apoptosis. The binding of cytokines and growth factors to cell surface receptors activates intracellular signal transduction cascades that promote cell survival. Akt (PKB) phosphorylates BAD on Ser 136, and BAD has also been shown to be phosphorylated by PKA and p90RSK on Ser 112. Phosphorylated BAD binds to the 14-3-3 protein, thereby inhibiting its interaction with Bcl-2 and Bcl-xL by its cytosolic sequestration. The anti-apoptotic proteins Bcl-2 and Bcl-xL are then free to interact with Apaf-1 and BID, thus promoting cell survival.

Figure 1: BAD signaling within the cell

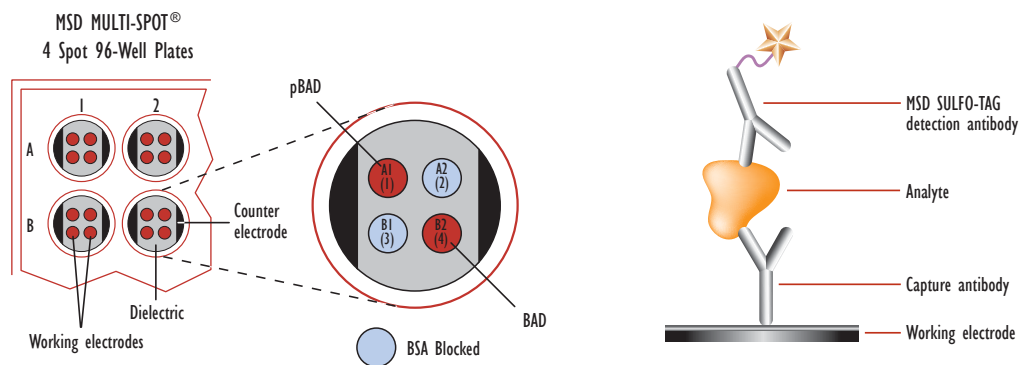


Principle of the Assay

principle of the assay

MSD® biomarker detection assays provide a rapid and convenient method for measuring the total and phosphorylated levels of protein targets within a single small-volume sample. These assays are available in both singleplex and multiplex formats. In a singleplex assay, an antibody for a specific protein target is coated on one electrode (or "spot") per well. In a multiplex assay, an array of capture antibodies against different targets is patterned on distinct spots in the same well. Our duplex assay for phospho- and total BAD uses sandwich immunoassays (Figure 2). MSD provides a plate that has been pre-coated with the capture antibodies (antibodies for phospho-(Ser 112) and total BAD). The user adds the sample and a solution containing the labeled detection antibody (anti-total BAD antibody labeled with an electrochemiluminescent compound, MSD SULFO-TAG™ label) over the course of one or more incubation periods. Phosphorylated and unphosphorylated BAD in the sample bind to the capture antibodies immobilized on the working electrode surface; recruitment of the labeled detection antibody by bound BAD completes the sandwich. The user adds an MSD Read Buffer that provides the appropriate chemical environment for ECL and loads the plate into an MSD SECTOR® Imager for analysis. Inside the SECTOR Imager, a voltage applied to the plate electrodes causes the labels bound to the electrode surface to emit light. The instrument measures intensity of the emitted light to afford a quantitative measure of the amount of phosphorylated and total BAD present in the sample.

Figure 2: MSD MULTI-SPOT Assay



The SECTOR Imager data file will identify spots according to their well location, not by the coated capture antibody name.

Diagram of Completed Reaction on Spot 1

Reagents Supplied

reagents supplied

| Material | Storage |
|---|---------|
| Blocker A (dry powder) | RT |
| Read Buffer T (with surfactant), 4X | RT |
| MULTI-SPOT 96-well Plate Phospho (Ser 112)/ Total BAD | 2-8°C |
| SULFO-TAG Detection Antibody (50X) | 2-8°C |
| Tris Wash Buffer (10X) | 2-8°C |
| Tris Lysis Buffer (1X) | 2-8°C |
| Phosphatase Inhibitor I (100X) | 2-8°C |
| Phosphatase Inhibitor II (100X) | 2-8°C |
| Protease Inhibitor Solution (50X) | ≤-10°C |

IV Required Materials and Equipment- not supplied

Required Materials and Equipment - not supplied

- Deionized water for diluting concentrated buffers
- 250 mL bottle (1)
- 50 mL tube (2)
- 15 mL tube (1)
- Adhesive plate seals
- Microtiter plate shaker
- Microcentrifuge tubes (various) for preparing serial dilutions of samples
- Appropriate liquid handling equipment for desired throughput, capable of accurately dispensing 25 and 150 μ L into a 96-well microtiter plate
- Plate washing equipment: automated plate washer, or other efficient multi-channel pipetting equipment

V Optional Material- not supplied

Optional Materials - not supplied

- Phospho-BAD Whole Cell Lysate Set (C11CC-1)

Safe laboratory practices and personal protective equipment such as gloves, safety glasses, and a lab coat should be used at all times during the handling of all kit components. All hazardous samples should be handled and disposed of properly, in accordance with local, state, and federal guidelines.

VII Reagent Preparation

r e a g e n t p r e p a r a t i o n

1. Tris Wash Buffer:

Dilute 10X stock of Tris Wash Buffer provided with the MSD kit to 1X as shown below. Tris Wash Buffer (1X) will be used throughout the assay to make additional reagents and wash plates. Approximately 250 mL per plate is required - more if using an automatic plate washer.

Combine (per plate):

- 25 mL 10X Tris Wash Buffer
- 225 mL deionized water

Excess Tris Wash Buffer may be stored at room temperature in a tightly sealed container for later use.

2. Blocking Solution:

Combine (per plate):

- 600 mg Blocker A (dry powder)
- 20 mL 1X Tris Wash Buffer

3. Antibody Dilution Buffer:

Combine (per plate):

- 1 mL Blocking Solution
- 2 mL Tris Wash Buffer

Set aside on ice.

4. Complete Lysis Buffer:

To 10 mL of Tris Lysis Buffer provided with the MSD kit, add the following supplemental materials to prepare the Complete Lysis Buffer (sufficient for 2-3 plates):

- 200 μ L Protease Inhibitor Solution (50X stock)
- 100 μ L Phosphatase Inhibitor I (100X stock)
- 100 μ L Phosphatase Inhibitor II (100X stock)

Keep Complete Lysis Buffer on ice until use.

5. Detection Antibody Solution:

Combine (per plate):

- 2.94 mL Antibody Dilution Buffer
- 60 μ L 50X SULFO-TAG Anti-Total BAD Antibody (1X final concentration)

6. 1X Read Buffer:

Combine (per plate):

- 15 mL deionized water
- 5 mL 4X Read Buffer T, with surfactant

VIII Sample Preparation and Storage

sample preparation and storage

This cell lysis protocol is provided as a reference. Specific cell types or targets may benefit from alternative buffer components or techniques, depending upon the particular research application. Most lysis buffers are compatible with MSD MULTI-SPOT plates, although high concentrations of denaturing detergents (>0.1 %) and reducing agents (DTT >1 mM) should be avoided. Please contact MSD Customer Support with any questions regarding lysate preparation options.

All manipulations should be performed on ice. Prepare Complete Lysis Buffer. The amount of Complete Lysis Buffer required will vary depending on scale of preparation and type of cells. Larger cells (e.g. NIH3T3, HeLa) should be lysed at concentrations of $1-5 \times 10^6$ cells per mL of lysis buffer. Smaller cells (e.g. Jurkat) should be lysed at concentrations of $1-5 \times 10^7$ cells per mL of lysis buffer.

Cells should be prepared as desired to activate target protein.

Phosphate Buffered Saline (PBS) should be chilled ice-cold.

Suspension cells

Pellet cells by centrifugation at $500 \times g$ at 4°C for 3 minutes. Discard supernatant and wash the pellet once with cold PBS. Pellet the cells again, discard supernatant, and resuspend in Complete Lysis Buffer at $1-5 \times 10^7$ cells per mL. Incubate on ice for 30 minutes. A shorter incubation time of 15 minutes may be adequate for many targets. Clear cellular debris from the lysate by centrifugation greater than or equal to $10,000 \times g$, at 4°C for 10 minutes. Discard the pellet and determine protein concentration in the lysate using a detergent compatible protein assay such as BCA. Unused lysates should be aliquoted and quickly frozen in a dry ice-ethanol bath and stored at -80°C .

Adherent cells

All volumes are determined for cells plated in 15 cm dishes. Remove media from the plates and wash cells one time with 5 mL cold PBS. Add 2 mL PBS to the plates and scrape the cells from the surface of the dish and transfer into 15 mL conical tubes. Pellet the cells by centrifugation at 500 x g for 3 minutes at 4°C. Discard supernatant and resuspend cells in 0.5-2 mL of Complete Lysis Buffer per dish. Incubate on ice for 30 minutes. A shorter incubation time of 15 minutes may be adequate for many targets. Clear cellular debris from the lysate by centrifugation greater than or equal to 10,000 x g, at 4°C for 10 minutes. Discard the pellet and determine protein concentration in the lysate using a detergent compatible protein assay such as BCA. Unused lysates should be aliquoted and quickly frozen in a dry ice-ethanol bath and stored at -80°C.

Refer to Appendix I (page 16) for cell lysate preparation protocol modifications that accommodate the use of 96-well culture plates.

IX Protocol

protocol

**Read entire protocol prior to beginning assay.
Please contact MSD Customer Support with any questions.**

The following protocol describes the most conservative approach to achieving optimal results with the Phospho (Ser 112)/ Total BAD Assay. The entire assay, including plate analysis on the MSD reader, can be completed in 3.5 hours. Once desired results are achieved, the protocol can be streamlined to eliminate multiple incubation and wash steps. Samples may be prepared for testing in the manner outlined in Sample Preparation and Storage, section VIII.

STEP 1: Block Plate and Prepare Samples.

- a) Add 150 µL/well of Blocking Solution A.
- b) Incubate with shaking for 1 hour at room temperature. Prepare Complete Lysis Buffer and dilute samples during this time.

Note: Samples, including cell lysates, etc., may be used neat or after dilution.

- MSD plates are compatible with most sample matrices. Avoid reagents that will denature the capture antibodies (e. g. high concentrations of reducing agents such as DTT should be avoided, and also SDS and other ionic detergents should be 0.1% or less in the sample applied to the well).

Note:

Solutions containing MSD Blocker A should be stored at 4°C and discarded after 14 days.

Plates may also be blocked overnight at 4°C.

- Depending on the stability of the target in the matrix, additional protease and phosphatase inhibitors may be required in the matrix or diluent.
- If working with purified protein, only a few nanograms per well will generally provide a strong assay signal. Purified recombinant proteins may exhibit differences in both signal and background as compared to native proteins in cell lysates.
- Keep diluted samples on ice until use.

c) *Prepare positive and negative cell lysates: (if provided with kit)*

- 1) Thaw cell lysate samples on ice and dilute immediately before use. Keep on ice during all manipulations and discard all remaining thawed unused material.
 - 2) Dilute cell lysate in Complete Lysis Buffer to a final concentration of 0.8 $\mu\text{g}/\mu\text{L}$. This will deliver 20 $\mu\text{g}/\text{well}$ in 25 μL . A dilution series may also be prepared if desired.
 - 3) Keep diluted cell lysate on ice until use.
- d) Wash plates four times with at least 150 $\mu\text{L}/\text{well}$ Tris Wash Buffer.

STEP 2: Add Samples and Prepare Detection Antibody Solution.

- a) Dispense 25 $\mu\text{L}/\text{well}$ of samples.
- b) Incubate with shaking for 1 hour at room temperature. Prepare Detection Antibody Solution during this time.
- c) Wash plates four times with at least 150 $\mu\text{L}/\text{well}$ Tris Wash Buffer.

STEP 3: Add Detection Antibody.

- a) Add 25 $\mu\text{L}/\text{well}$ of Detection Antibody Solution.
- b) Incubate with shaking for 1 hour at room temperature. Prepare 1X Read Buffer during this time.
- c) Wash plates four times with at least 150 $\mu\text{L}/\text{well}$ Tris Wash Buffer.

Note:

Complete Lysis Buffer should be prepared the day of use and then discarded. It should be kept ice-cold during all experimental manipulations.

The sensitivity of MSD immunoassays rivals that of ELISAs and Western blots. The amount of sample required for a given assay will depend on the abundance of the analyte in the matrix and the affinities of the antibodies used.

Samples and standards cannot be serially diluted in the MSD plate. Use microcentrifuge tubes or a separate 96-well polypropylene plate to prepare dilutions.

Note:

Shaking a 96-well MSD MULTI-ARRAY® or MULTI-SPOT plate during an incubation step will accelerate capture at the working electrode.

The lysate sample incubation time provided is optimized for the use of MSD cell lysates. Samples from other sources may require a longer incubation.

Note:

Excess diluted Read Buffer may be kept in a tightly sealed container at room temperature for later use.

STEP 4: Read Plate.

- a) Carefully add 150 μ L/well of diluted Read Buffer T, with surfactant, avoiding the introduction of any bubbles.
- b) Analyze with SECTOR Imager:
 1. Double click on DISCOVERY WORKBENCH[®] icon on computer desktop (if not already open).
 2. Click the SECTOR Imager icon in upper left corner of screen (if not already open to plate reading screen).
 3. From the pull down menu select "Read From Barcode."
 4. If only reading one plate check "Return Plate to Input Stack".
 5. Check the box and enter number of plates to be read.
 6. Click the "Run" button.
 7. Check the box to export default data file.
 8. If desired, make selections to export a custom data file.
 9. Browse and select the location to export data files.
 10. Click OK to initiate the run.
 11. Data will be automatically saved in the software database and text versions of the requested data files exported to the folder designated.

Note:

Bubbles introduced during the Read Buffer addition will interfere with imaging of the plate and produce unreliable data.

Plates should be imaged within 5 minutes following the addition of Read Buffer. Due to the varying nature of each research application, assay stability should be investigated prior to allowing plates to sit with Read Buffer for extended periods.

An all-inclusive indelible copy of the data and associated instrument information will be saved on the internal database, regardless of data file export selection. Additional copies of the data can be exported in any layout at a later time using this database. Consult the instrument user manual for more information.

Note: Once the assay is fully developed and producing desired results, the protocol listed above can be streamlined by combining the lysate and antibody incubation steps and removing the corresponding washes. A checklist summary protocol is seen below with a sample assay plate layout.

- Add blocking solution and incubate.
- Wash.
- Add samples and incubate.
- Wash.
- Add detection antibody solution and incubate.
- Wash.
- Add Read Buffer and analyze plate.

| Cell Lysate (µg) | | Control Samples | | | | | | | | Test Samples | | | |
|---------------------|---|-----------------|---|---|---|---------------|---|---|---|--------------|----|----|----|
| | | pBAD Positive | | | | pBAD Negative | | | | Unknown | | | |
| | | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 |
| 0 | A | | | | | | | | | | | | |
| | B | | | | | | | | | | | | |
| 5 | C | | | | | | | | | | | | |
| | D | | | | | | | | | | | | |
| 10 | E | | | | | | | | | | | | |
| | F | | | | | | | | | | | | |
| 20 | G | | | | | | | | | | | | |
| | H | | | | | | | | | | | | |

Figure 3: Sample assay plate layout. MSD Control Cell Lysates expected to produce signal for phosphorylated and total BAD are shaded in yellow and total BAD only are shaded in green.

X Calculation of Results

calculation of results

The percent phosphoprotein in a sample can be calculated using independent MSD total and phospho singleplex phosphoprotein assays or MSD total/phospho multiplex phosphoprotein assays. MSD has optimized the amount of capture antibody used in multiplex assays to account for differences in binding affinities between anti-phospho and anti-total capture antibodies, as multiple capture antibodies may compete for the same analyte in a multiplex assay format.

INDEPENDENT ASSAY FORMAT: Anti-Total Singleplex and Anti-Phospho Singleplex using the same detection antibody

$$\% \text{ Phosphoprotein} = (\text{Phospho signal} / \text{Total signal}) * 100$$

MULTIPLEX ASSAY FORMAT: Anti-Total and Anti-Phospho Assay in the same well

$$\% \text{ Phosphoprotein} = ((2 * \text{Phospho signal}) / (\text{Phospho signal} + \text{Total signal})) * 100$$

Note: The numerator is 2X the phospho signal because the phosphorylated species is captured by both the phospho-specific and the phosphorylation state-independent capture antibodies, and the denominator is "phospho + total" signal because the true total is all of the material captured on both spots.

The following points should be noted when calculating percent phosphoprotein using MSD assays:

- The capture antibodies used in MSD multiplex assays may differ in their weak binding to abundant proteins in the test lysate, therefore each assay in the well may not be linear over the same concentration range.
- Different protein targets may vary greatly in abundance within a particular sample, therefore the establishment of a linear range for each target is recommended.

Sample Data and Calculations for Each Assay Format:

Jurkat cell lysates were untreated or treated with the inhibitor LY294002. Capture antibodies used for plate coating are indicated.

For demonstration purposes, calculations for percent phosphoprotein are shown here using data from MSD MULTI-SPOT Phospho-Akt, Total Akt, and Phospho/Total Akt Assays.

INDEPENDENT ASSAYS

| Phospho-Akt Capture Antibody | | | | | | | | |
|------------------------------|---------------|--------|-----|---------------|--------|-----|-------|------|
| Jurkat lysates | pAkt Positive | | | pAkt Negative | | | P-N | P/N |
| (μ g) | Ave | StdDev | %CV | Ave | StdDev | %CV | | |
| 0 | 121 | 6 | 5 | 104 | 1 | 1 | | |
| 0.3 | 856 | 66 | 8 | 144 | 14 | 10 | 712 | 5.9 |
| 0.6 | 1370 | 87 | 6 | 169 | 13 | 8 | 1201 | 8.1 |
| 1.3 | 2448 | 10 | 0 | 209 | 88 | 42 | 2239 | 11.7 |
| 2.5 | 4402 | 211 | 5 | 260 | 106 | 41 | 4142 | 16.9 |
| 5 | 8925 | 439 | 5 | 455 | 59 | 13 | 8470 | 19.6 |
| 10 | 16618 | 1653 | 10 | 752 | 83 | 11 | 15866 | 22.1 |
| 20 | 34590 | 3630 | 10 | 1103 | 64 | 6 | 33487 | 31.4 |

Figure 4: Phospho-Akt capture alone.

| Total Akt Capture Antibody | | | | | | | | |
|----------------------------|---------------|--------|-----|---------------|--------|-----|--------|-----|
| Jurkat lysates | pAkt Positive | | | pAkt Negative | | | P-N | P/N |
| (μ g) | Ave | StdDev | %CV | Ave | StdDev | %CV | | |
| 0 | 243 | 47 | 19 | 218 | 8 | 4 | | |
| 0.3 | 1724 | 132 | 8 | 2588 | 165 | 6 | -865 | 0.7 |
| 0.6 | 3552 | 69 | 2 | 5489 | 453 | 8 | -1938 | 0.6 |
| 1.3 | 6530 | 115 | 2 | 9626 | 195 | 2 | -3097 | 0.7 |
| 2.5 | 11577 | 242 | 2 | 18481 | 75 | 0 | -6904 | 0.6 |
| 5 | 19820 | 1080 | 5 | 31255 | 77 | 0 | -11435 | 0.6 |
| 10 | 34149 | 474 | 1 | 50096 | 58 | 0 | -15948 | 0.7 |
| 20 | 45475 | 505 | 1 | 63923 | 1044 | 2 | -18448 | 0.7 |

Figure 5: Total Akt capture alone.

MULTIPLEX ASSAY

| Phospho-Akt Capture Antibody | | | | | | | | |
|------------------------------|---------------|--------|-----|---------------|--------|-----|-------|------|
| Jurkat lysates | pAkt Positive | | | pAkt Negative | | | P-N | P/N |
| (μ g) | Ave | StdDev | %CV | Ave | StdDev | %CV | | |
| 0 | 89 | 16 | 18 | 112 | 16 | 14 | | |
| 0.3 | 584 | 87 | 15 | 158 | 15 | 9 | 426 | 3.7 |
| 0.6 | 983 | 170 | 17 | 169 | 21 | 12 | 815 | 5.8 |
| 1.3 | 1373 | 208 | 15 | 269 | 12 | 4 | 1105 | 5.1 |
| 2.5 | 3124 | 742 | 24 | 294 | 28 | 9 | 2830 | 10.6 |
| 5 | 6087 | 3088 | 51 | 485 | 11 | 2 | 5602 | 12.6 |
| 10 | 15169 | 3718 | 25 | 732 | 60 | 8 | 14438 | 20.7 |
| 20 | 29258 | 4939 | 17 | 1173 | 189 | 16 | 28085 | 25.0 |

| Total Akt Capture Antibody | | | | | | | | |
|----------------------------|---------------|--------|-----|---------------|--------|-----|--------|-----|
| Jurkat lysates | pAkt Positive | | | pAkt Negative | | | P-N | P/N |
| (μ g) | Ave | StdDev | %CV | Ave | StdDev | %CV | | |
| 0 | 220 | 44 | 20 | 243 | 44 | 18 | | |
| 0.3 | 1705 | 255 | 15 | 3241 | 255 | 8 | -1537 | 0.5 |
| 0.6 | 3286 | 89 | 3 | 5853 | 89 | 2 | -2567 | 0.6 |
| 1.3 | 5629 | 1109 | 20 | 10390 | 1109 | 11 | -4762 | 0.5 |
| 2.5 | 11701 | 1050 | 9 | 19325 | 1050 | 5 | -7624 | 0.6 |
| 5 | 21189 | 3072 | 14 | 34547 | 3072 | 9 | -13359 | 0.6 |
| 10 | 33731 | 4094 | 12 | 52575 | 4094 | 8 | -18844 | 0.6 |
| 20 | 44540 | 14 | 0 | 71846 | 14 | 0 | -27307 | 0.6 |

Figure 6: Phospho Akt and total Akt in the same well.

CALCULATION OF PERCENT PHOSPHOPROTEIN BY EACH METHOD

| Jurkat lysates | % pAkt in pAkt Positive Lysates | | | % pAkt in pAkt Negative Lysates | | |
|----------------|---------------------------------|--|-----------|---------------------------------|--|----------|
| (μ g) | alone | | multi | alone | | multi |
| 0 | | | | | | |
| 0.3 | 50 | | 51 | 6 | | 9 |
| 0.6 | 39 | | 46 | 3 | | 6 |
| 1.3 | 37 | | 39 | 2 | | 5 |
| 2.5 | 38 | | 42 | 1 | | 3 |
| 5 | 45 | | 45 | 1 | | 3 |
| 10 | 49 | | 62 | 2 | | 3 |
| 20 | 76 | | 79 | 2 | | 3 |

Figure 7: Independent and multiplex calculation of percent phosphoprotein in a cell lysate titration. Untreated Jurkat cell lysate is expected to express phosphorylated Akt. Treatment with LY294002 inhibitor is expected to inhibit phosphorylated Akt production but not affect total Akt levels.

XI Limitations of Procedure

limitations of procedure

The following points should be noted with the MULTI-SPOT Phospho (Ser 112)/ Total BAD Assay to maximize assay sensitivity and performance:

- A no-wash assay format may be employed, however lower sensitivity may be observed.
- All buffers containing phosphate should be avoided when detecting phosphoproteins.
- Due to the unstable nature of phosphoproteins, cell lysates should be thawed immediately prior to use and any remaining thawed material should be subsequently discarded.

XII Typical Data

typical data

Typical results for the MULTI-SPOT Phospho (Ser 112)/ Total BAD Assay are illustrated in Figures 8-10 below. Please note that these signal and ratio values are provided for demonstration only and individual results may vary depending upon samples tested.

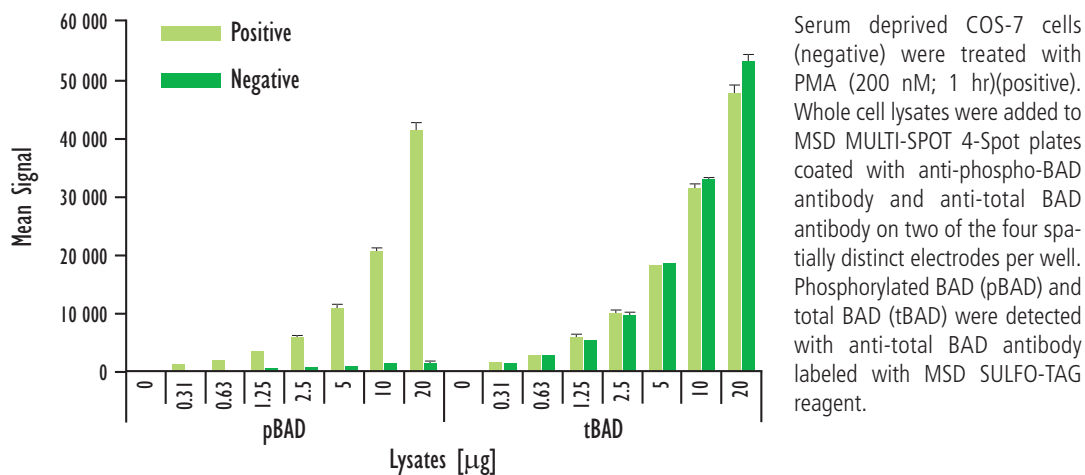
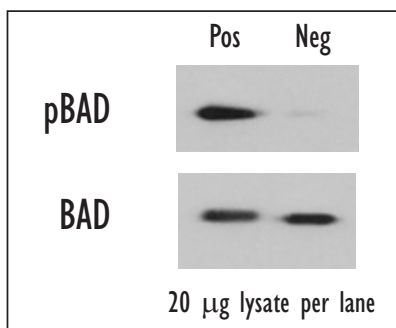


Figure 8: Sample data generated with the MSD MULTI-SPOT Phospho (Ser 112)/ Total BAD Assay. Increased signal for phosphorylated BAD was observed with only pBAD positive cell lysates as total BAD signal increased throughout the titration of both pBAD positive and negative cell lysates. The MSD MULTI-SPOT Phospho (Ser 112)/ Total BAD Assay provides a quantitative measure of the data obtained with the traditional Western blot.



Lysates were prepared as described in Section VIII, Sample Preparation and Storage, from COS-7 cells treated as outlined in Figure 8. Western blot analysis of each lysate type was performed with phospho (Ser 112)-specific and total BAD antibodies.

Figure 9: Western Blot analysis of cell lysate. The signal generated in the MULTI-SPOT Assay is consistent with the qualitative Western blot results.

| | Lysates (μ g) | Positive | | | Negative | | | P/N |
|------|--------------------|----------|--------|-----|----------|--------|-----|------|
| | | Average | StdDev | %CV | Average | StdDev | %CV | |
| pBAD | 0 | 51 | 4 | 8 | 50 | 5 | 10 | |
| | 0.31 | 1195 | 42 | 3 | 270 | 15 | 6 | 4.4 |
| | 0.63 | 2049 | 15 | 1 | 378 | 6 | 2 | 5.4 |
| | 1.25 | 3579 | 4 | 0 | 497 | 11 | 2 | 7.2 |
| | 2.5 | 6071 | 85 | 1 | 682 | 12 | 2 | 8.9 |
| | 5 | 11003 | 482 | 4 | 1129 | 36 | 3 | 9.7 |
| | 10 | 20594 | 629 | 3 | 1365 | 93 | 7 | 15.1 |
| | 20 | 41197 | 1304 | 3 | 1689 | 108 | 6 | 24.4 |
| tBAD | 0 | 148 | 8 | 5 | 147 | 5 | 3 | |
| | 0.31 | 1582 | 31 | 2 | 1465 | 9 | 1 | 1.1 |
| | 0.63 | 2980 | 40 | 1 | 2787 | 59 | 2 | 1.1 |
| | 1.25 | 5975 | 431 | 7 | 5316 | 8 | 0 | 1.1 |
| | 2.5 | 10153 | 449 | 4 | 9800 | 388 | 4 | 1.0 |
| | 5 | 18207 | 143 | 1 | 18523 | 40 | 0 | 1.0 |
| | 10 | 31206 | 823 | 3 | 32876 | 408 | 1 | 0.9 |
| | 20 | 47703 | 1269 | 3 | 53211 | 1141 | 2 | 0.9 |

Figure 10: Lysate titration data for pBAD positive and negative COS-7 cell lysates using the MSD MULTI-SPOT Phospho (Ser 112)/ Total BAD Assay.

XIII References

references

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MULTI-ARRAY Phospho-BAD (Ser 112) Assay

MULTI-ARRAY Total BAD Assay

Phospho- and total BAD antibodies can be used in custom phosphorylated and total protein multiplex assay panels designed to meet specific research needs. Please contact MSD Customer Service for more information.

MSD Customer Service

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96-well Culture Plate Modifications

Successful adaptation to a 96-well culture format is cell type and target-dependent. The number of cells to be plated per well should be determined for each cell type. General recommended plating concentrations for adherent cells range from 1×10^4 - 5×10^4 cells per well, and approximately 2×10^6 cells per mL (50-75 μ L per well) for suspension cells. These numbers are provided as a guide and the optimal concentrations will vary depending upon cell line used.

Suspension cells

For flat bottom plates, experiments should be designed such that the final volume per well is 50-75 μ L. Perform cell lysis using a 4X Complete Lysis Buffer concentrate, supplemented with protease and phosphatase inhibitors at 4X concentrations. Add 4X Complete Lysis Buffer directly to cells in the growth medium for a final 1X concentration in the well.

NOTE: With some effort, a 10X Complete Lysis Buffer can also be prepared.

(For conical microwell plates, perform lysis by pelleting the cells, removing most of the growth medium and adding a constant amount of 1X Complete Lysis Buffer.)

Adherent cells

Plate cells on biologically treated tissue culture ware (such as BD BioCoat™ Cellware (Becton, Dickinson and Company, Franklin Lakes, NJ)) to reduce variability due to cells lost as growth medium is removed. Treat cells as desired. Gently aspirate growth medium from microwell plate. A PBS wash step is not required and can introduce variability. Add 50-100 μ L 1X Complete Lysis Buffer per well.

Cell lysis time should be determined by the end user. Some targets are immediately available for detection, while others may require an incubation step at room temperature, 4°C, or on ice with gentle agitation.

Carefully pipet cell lysate onto prepared capture plate and proceed with assay protocol. *It is important to transfer a constant volume and avoid pipeting too vigorously, as the introduction of air bubbles may result. (Targets can be captured from a volume greater than 25 μ L.)*

