

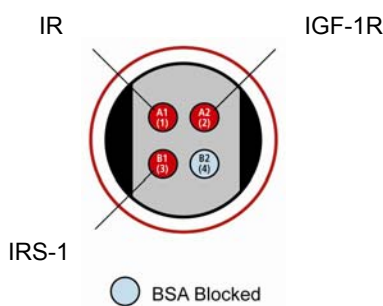
MSD[®] 96-Well MULTI-SPOT[®] Insulin Signaling Panel (Phosphoprotein)

The following assay protocol has been optimized for quantifying phosphorylated Insulin Signaling proteins (IGF-1R, IR and IRS-1) in whole cell lysate.

Storage

Materials Included

<input type="checkbox"/> Read Buffer T (with surfactant), 4X	RT
<input type="checkbox"/> Blocker A	RT
<input type="checkbox"/> MULTI-SPOT 96-well 4 Spot Insulin Signaling Panel Plate(s)	2-8°C
<input type="checkbox"/> SULFO-TAG [™] Anti-PY20 Antibody (50X)	2-8°C
<input type="checkbox"/> Tris Wash Buffer (10X)	2-8°C
<input type="checkbox"/> Tris Lysis Buffer (1X)	2-8°C
<input type="checkbox"/> Phosphatase Inhibitor I (100X)	2-8°C
<input type="checkbox"/> Phosphatase Inhibitor II (100X)	2-8°C
<input type="checkbox"/> Protease Inhibitor Solution (50X)	≤-10°C



The SECTOR[®] Imager data file will identify spots according to their well location, not by the coated capture antibody name.

FOR RESEARCH USE ONLY.
NOT FOR USE IN DIAGNOSTIC OR THERAPEUTIC PROCEDURES.



Notes:

Other Materials & Equipment (not supplied)

- Deionized water for diluting Wash Buffer and Read Buffer
- One 250 mL bottle
- Two 50 mL tube
- One 15 mL tube
- Adhesive plate seals
- Microtiter plate shaker
- Various microcentrifuge tubes for making serial dilutions of lysates (if desired)
- Automated plate washer or other efficient multi-channel pipetting equipment for washing 96-well plates
- Appropriate liquid handling equipment for desired throughput that must accurately dispense 25 μ L and 150 μ L into a 96-well micro plate

Read the entire detailed instructions before beginning work.

Protocol at a Glance

The following protocol describes the most conservative approach toward achieving highly sensitive results using MSD technology to quantify phosphoproteins. The protocol can be completed in approximately 3 to 3 1/2 hours if each reagent is prepared during the preceding incubation. All reagents with the exception of diluted lysates can also be prepared ahead of time. This lengthens the overall time required for the assay but frees up time during incubation steps.

Once desired results are achieved, the protocol can be streamlined to eliminate multiple incubation and wash steps to increase throughput.

1. Add Blocking Solution, incubate 1 hour, wash.
2. Add lysate, incubate 1 hour at room temperature, wash.
3. Add Detection Antibody, incubate 1 hour, wash.
4. Add Read Buffer and analyze plate.

A larger amount of Tris Wash Buffer may be prepared at once and stored at room temperature for later use.

Detailed Instructions

Prepare a stock of 1X Tris Wash Buffer:

- a) 1X Tris Wash Buffer will be used throughout the assay to make other reagents as well as wash plates. Approximately 250 mL per plate is required – more if using an automatic plate washer.
- b) In a 250 mL bottle combine:
 - 25 mL 10X Tris Wash Buffer
 - 225 mL deionized water



Notes:

Solutions containing Blocker A should be kept at 4°C and discarded after 14 days.

Prepare Blocking Solution-A:

- a) Prepare 20 mL per plate.
- b) In a 50 ml tube combine:
 - 20 mL 1X Tris Wash Buffer
 - Blocker A 600mg (3%)

Prepare Antibody Dilution Buffer:

- a) Prepare 3 mL per plate.
- b) In a 15 mL tube combine:
 - 1 mL Blocking Solution-A
 - 2 mL 1X Tris Wash Buffer

Begin with a MULTI-SPOT Insulin Signaling Panel Plate.
No pre-treatment is necessary.

STEP 1

Add 150 µL/well of Blocking Solution.

Plates may also be blocked overnight at 4°C.

Incubate with shaking at room temperature for 1 hour. Prepare Complete Tris Lysis Buffer, and dilute cell lysates during this time.

Shaking a 96-well MSD MULTI-ARRAY® or MULTI-SPOT plate accelerates capture at the working electrode.

Prepare Complete Tris Lysis Buffer:

- a) It is important that all of the reagents (including the Tris Lysis Buffer) are room temperature before mixing. To 10 mL of Tris Lysis Buffer, add the following:
 - 100 µL Phosphatase Inhibitor I (100X stock)
 - 100 µL Phosphatase Inhibitor II (100X stock)
 - 200 µL Protease Inhibitor Solution (50X stock)
- b) Keep Complete Tris Lysis Buffer on ice until use.

Complete Tris Lysis Buffer should be made each day of experimentation.

The Complete Tris Lysis Buffer should be ice cold before use.

Prepare samples or positive and negative cell lysates:

(Note: Recommendations for cell lysate handling are provided; however, the suggested concentrations listed below may need to be adjusted depending upon specific samples tested.)

- a) Thaw cell lysate samples on ice and dilute immediately before use. Keep on ice during all manipulations and discard all remaining thawed unused material.
- b) Dilute positive and negative cell lysates in Complete Tris Lysis Buffer to a final concentration of 0.8 µg/µL. This will deliver 20 µg/well in 25 µL. A dilution series may also be prepared if desired.

Wash plates three times with Wash Buffer.



Notes:

STEP 2 Dispense 25 μ L/well of diluted lysates.

Incubate with shaking for 1 hour at room temperature. Prepare Detection Antibody Solution during this time.

Prepare Detection Antibody Solution:

- a) Prepare 3.0 mL per plate.
- b) In a 15 mL tube combine:
 - a. 2.94 mL cold Antibody Dilution Buffer
 - b. 60 μ L 50X Anti-PY20 Antibody
(Final concentration: 1X)

Wash plates three times with Wash Buffer.

STEP 3 Add 25 μ L/well of Detection Antibody Solution.

Incubate with shaking at room temperature for 1 hour. Prepare Read Buffer during this time.

Dilute Read Buffer:

- a) Approximately 20 mL per plate is required.
- b) Dilute 4X Read Buffer T (with surfactant) to 1X with deionized water.

Wash plates three times with Wash Buffer.

Diluted Read Buffer may be kept in a tightly sealed container at room temperature for later use.

Plates can be imaged immediately following the addition of read buffer.

STEP 4 Add 150 μ L/well of diluted Read Buffer T (with surfactant).

Analyze with SECTOR Imager plate reader.

Bubbles in the Read Buffer will interfere with reliable imaging of the plate if carried into the wells.

