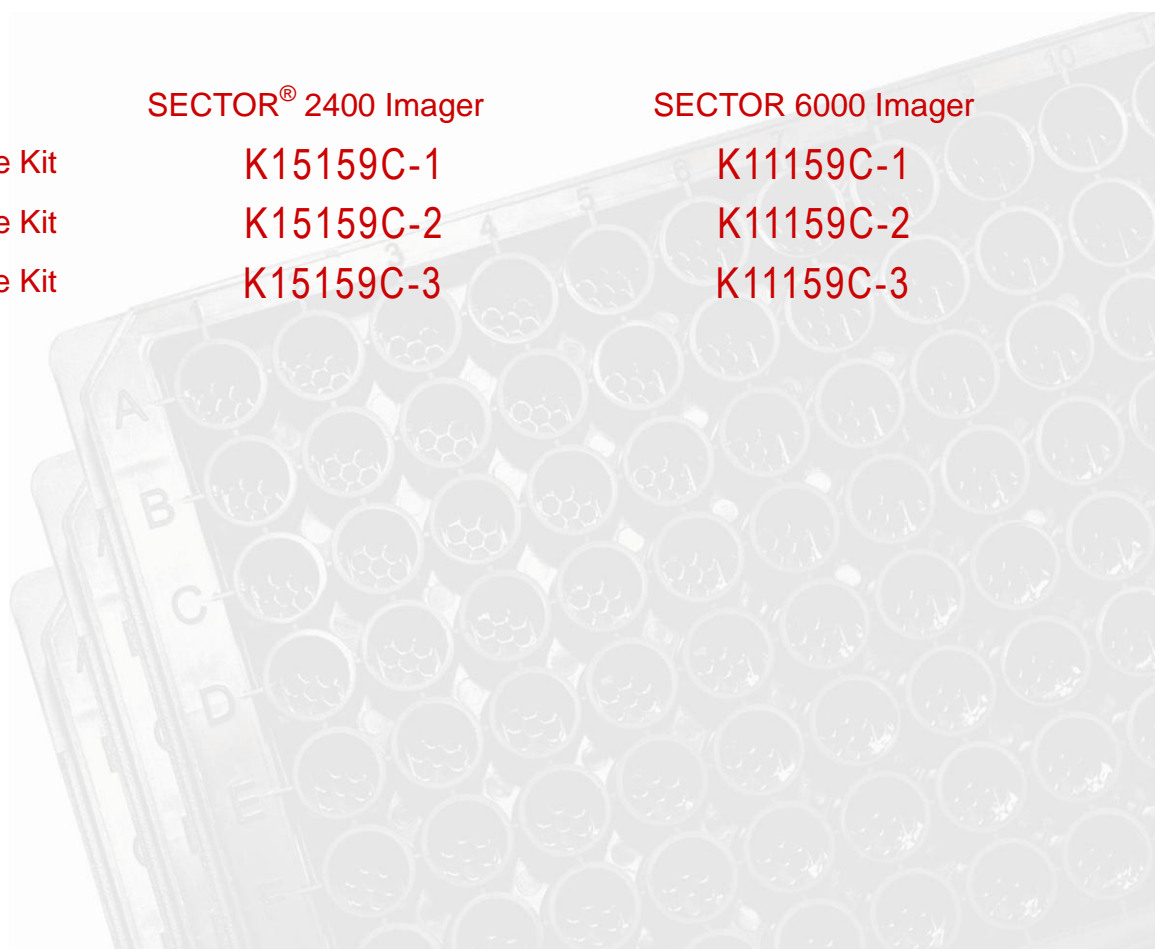


Meso Scale Discovery[®]

MULTI-SPOT[®] Assay System

Mouse/Rat GLP-1 (7-36)amide, Insulin,
Glucagon Assay Kit

	SECTOR [®] 2400 Imager	SECTOR 6000 Imager
1-Plate Kit	K15159C-1	K11159C-1
5-Plate Kit	K15159C-2	K11159C-2
20-Plate Kit	K15159C-3	K11159C-3



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MSD Metabolic Assays

Mouse/Rat GLP-1 (7-36)amide, Insulin, Glucagon Assay Kit

This package insert must be read in its entirety before using this product.

FOR RESEARCH USE ONLY.

NOT FOR USE IN DIAGNOSTIC OR THERAPEUTIC PROCEDURES.

Meso Scale Discovery

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Table of Contents

table of contents

Introduction.....	4
Principle of the Assay	5
Reagents Supplied	6
Required Material and Equipment – not supplied	6
Safety	7
Reagent Preparation	7
Assay Protocol.....	9
Analysis of Results	9
Typical Standard Curve	10
Sensitivity	11
Spike Recovery	11
Linearity	11
Cross-Reactivity	12
Kit Components	12
References	14
Summary Protocol	15
Plate Diagrams	17

Ordering Information

ordering information

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Introduction

introduction

Glucagon-like peptide-1 (GLP-1) is a 3.5 kD protein hormone produced in intestinal L cells and is associated with lowering blood glucose levels. By activation of different physiological systems, it plays roles in gastric emptying upon intake of nutrients, the regulation of short-term feeding behavior, the promotion of glucose-dependent insulin secretion and insulin biosynthesis, and also the inhibition of glucagon secretion. The cleaved peptides, commonly referred to as GLP-1 (7-36)amide and GLP-1 (7-37), are the biologically active forms of GLP-1. *In vivo*, the amidated form is rapidly degraded by dipeptidyl peptidase IV (DPP IV). Since GLP-1, in its bioactive form, plays a crucial role in blood glucose regulation, GLP-1 mimetics and inhibitors of DPP IV are currently being evaluated as potential drug candidates in treatment of diabetes. MSD offers a comprehensive array of GLP-1 assays that measure both the active and total GLP-1 protein using detection antibodies specific for the C-terminal, 36th and/or 37th amino acids.

Insulin is a 51-residue peptide hormone that is produced in the pancreas by β -cells of the islets of Langerhans. Insulin is involved in the regulation of carbohydrate, fat and protein metabolism. Lowered levels of insulin cause liver cells to convert glycogen back to glucose and secrete it into the blood. Insulin also has an effect on small vessel muscle tone, storage and release of (fat) triglycerides and cellular uptake of amino acids and electrolytes. Type 1 diabetes results when the β -cells are destroyed and no longer producing insulin resulting in high glucose levels in the blood. Patients with type 1 diabetes depend on exogenous insulin for their survival because of an absolute deficiency of the hormone; patients with type 2 diabetes have either relatively low insulin production or insulin resistance or both.

Glucagon is a 29-residue polypeptide hormone that is produced in the pancreas by the α -cells of the islets of Langerhans. Glucagon is involved in maintaining normal levels of glucose in the blood by acting on liver glycogen, converting it to glucose. Glucagon is a stimulator of hepatic glycogenolysis, gluconeogenesis, and ketogenesis which are antagonistic effects to those of insulin action, resulting in increased blood glucose levels. Glucagon receptors have been found in liver, kidney, intestinal smooth muscle, brain and adipose tissue.

Principle of the Assay

principle of the assay

MSD[®] metabolic assays provide a rapid and convenient method for measuring the levels of protein targets within a single small-volume sample. The assays are available in both singleplex and multiplex formats. In a singleplex assay, an antibody for a specific protein target is coated on one electrode (or “spot”) per well. In a multiplex assay, an array of capture antibodies against different targets is patterned on distinct spots in the same well. Our Mouse/Rat GLP-1 (7-36)amide, Insulin, Glucagon Assay detects all amidated forms of GLP-1, insulin and glucagon in a multiplexed sandwich immunoassay (Figure 1). MSD provides a plate that has been pre-coated with GLP-1 total, insulin and glucagon capture antibodies. The user adds the sample and a solution containing the labeled detection antibodies—anti-GLP-1 (7-36)amide, anti-insulin and anti-glucagon labeled with an electrochemiluminescent compound, MSD SULFO-TAG[™] label—over the course of one or more incubation periods. GLP-1 (7-36)amide, insulin and glucagon in the sample bind to their specific capture antibody immobilized on the working electrode surface; recruitment of each labeled detection antibody by bound analyte completes the sandwich. The user adds an MSD read buffer that provides the appropriate chemical environment for electrochemiluminescence and loads the plate into an MSD SECTOR instrument for analysis. Inside the SECTOR instrument, a voltage applied to the plate electrodes causes the labels bound to the electrode surface to emit light. The instrument measures intensity of emitted light to afford a quantitative measure of all amidated forms of GLP-1 {isoforms: (1-36), (7-36) and (9-36)} (Figure 2), insulin and glucagon present in the sample.

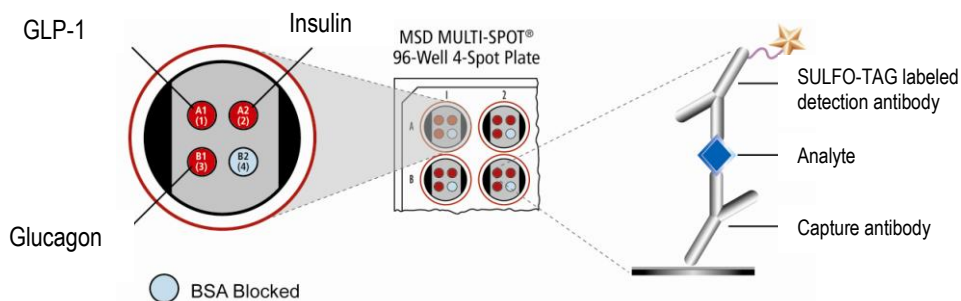


Figure 1. Sandwich immunoassay on MSD platform. The numbering convention for the different spots is maintained in the software visualization tools, on the plate packaging, and in the data files. Any spot that is not coated with a specific capture antibody is blocked with BSA to reduce non-specific binding to that spot. A unique bar code label on each plate allows complete traceability back to MSD manufacturing records.

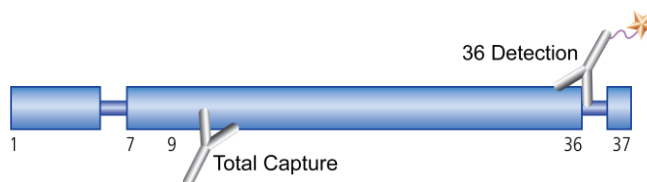


Figure 2. Schematic of the antibodies recognition sites for the GLP-1 (7-36)amide Assay on GLP-1 protein amino acids 1-37.

Reagents Supplied

reagents supplied

Product Description	Storage	Quantity per Kit		
		K15159C-1 K11159C-1	K15159C-2 K11159C-2	K15159C-3 K11159C-3
MULTI-SPOT 96-well M/R GLP-1, Insulin, Glucagon Plate(s) N45159A-1 (K15159C) N41159A-1 (K11159C)	2-8°C	1 plate	5 plates	20 plates
SULFO-TAG Anti-GLP-1 (7-36)amide Antibody ¹ (100X)	2-8°C	1 vial (40 µL)	1 vial (200 µL)	4 vials (200 µL ea)
SULFO-TAG Anti-m/r Insulin Antibody ¹ (100X)	2-8°C	1 vial (40 µL)	1 vial (200 µL)	4 vials (200 µL ea)
SULFO-TAG Anti-Glucagon Antibody ¹ (100X)	2-8°C	1 vial (40 µL)	1 vial (200 µL)	4 vials (200 µL ea)
GLP-1 (7-36)amide Calibrator 1 µg/mL	≤-70°C	1 vial (15 µL)	5 vials (15 µL ea)	20 vials (15 µL ea)
Insulin Calibrator 5 µg/mL	≤-70°C	1 vial (15 µL)	5 vials (15 µL ea)	20 vials (15 µL ea)
Glucagon Calibrator 1 µg/mL	≤-70°C	1 vial (15 µL)	5 vials (15 µL ea)	20 vials (15 µL ea)
Blocker A kit R93AA-2 (250 mL)	RT	1 bottle (250 mL)	1 bottle (250 mL)	4 bottles (250 mL ea)
Aprotinin (200,000 KIU/mL)	2-8°C	1 vial (50 µL)	1 vial (250 µL)	4 vials (250 µL ea)
Blocker D-B (10%)	≤-10°C	1 vial (0.25 mL)	1 vial (1.2 mL)	4 vials (1.2 mL ea)
Diluent 17	≤-10°C	1 bottle (30 mL)	2 bottles (30 mL ea)	6 bottles (30 mL ea)
Diluent 100 R50AA-4 (50 mL)	2-8°C	1 bottle (50 mL)	1 bottle (50 mL)	2 bottles (50 mL ea)
Read Buffer T (with surfactant), 4X R92TC-3 (50 mL) R92TC-2 (200 mL)	RT	1 bottle (50 mL)	1 bottle (50 mL)	1 bottle (200 mL)

Required Materials and Equipment - not supplied

required materials and equipment — not supplied

- Deionized water for diluting concentrated buffers
- 50 mL tubes for reagent preparation
- 15 mL tubes for reagent preparation
- Microcentrifuge tubes for preparing serial dilutions
- Phosphate buffered saline plus 0.05% Tween-20 (PBS-T) for plate washing
- Appropriate liquid handling equipment for desired throughput, capable of dispensing 10 to 150 µL into a 96-well microtiter plate
- Plate washing equipment: automated plate washer or multichannel pipette
- Adhesive plate seals
- Microtiter plate shaker

¹ Some SULFO-TAG labeled detection antibodies may be light-sensitive, so they should be stored in the dark.

V Safety

s a f e t y

Safe laboratory practices and personal protective equipment such as gloves, safety glasses, and lab coats should be used at all times during the handling of all kit components. All hazardous samples should be handled and disposed of properly, in accordance with local, state, and federal guidelines.

VI Reagent Preparation

r e a g e n t p r e p a r a t i o n

Bring all plates and diluents to room temperature.

Blocker D-B can tolerate up to 5 freeze-thaw cycles. Alternatively, an aliquot of the blocker can be stored at 2-8°C for up to 1 month.

Important: Upon first thaw, separate Diluent 17 into aliquots appropriate to the size of your assay needs. This diluent can go through up to three freeze-thaw cycles without significantly affecting the performance of the assay.

Prepare Blocker A Solution

Follow instructions included with the Blocker A Kit.

Prepare Metabolic Assay Working Solution

In a 15 mL tube combine (per plate):

- 40 µL of Aprotinin
- 7960 µL of Diluent 17

Important: Aprotinin should be added prior to use. The Metabolic Assay Working Solution should be kept on ice. Do not freeze the Metabolic Assay Working Solution for later use.

Prepare Calibrator and Control Solutions

The stock Calibrator vials are supplied at 1 µg/mL for GLP-1 (7-36)amide and Glucagon and at 5 µg/mL for Insulin. For the assay, an 8-point standard curve is recommended with 3-fold serial dilution steps and a zero Calibrator. The table below shows the concentrations of the 8-point standard curve:

Standard	GLP-1 (7-36)amide conc. (pg/mL)	Insulin conc. (pg/mL)	Glucagon conc. (pg/mL)	Dilution Factor
Stock Cal. Vial	1000000	5000000	1000000	
STD-01	10000	50000	10000	100
STD-02	3333	16667	3333	3
STD-03	1111	5556	1111	3
STD-04	370	1852	370	3
STD-05	123	617	123	3
STD-06	41	206	41	3
STD-07	14	69	14	3
STD-08	0	0	0	n/a

To prepare this 8-point standard curve:

- 1) Prepare the highest Calibrator by adding 10 μL of 1 $\mu\text{g}/\text{mL}$ GLP-1 (7-36)amide, 10 μL of 1 $\mu\text{g}/\text{mL}$ Glucagon and 10 μL of 5 $\mu\text{g}/\text{mL}$ Insulin to 970 μL of Metabolic Assay Working Solution.
- 2) Prepare the next Calibrator by transferring 100 μL of the diluted Calibrator to 200 μL of Metabolic Assay Working Solution. Repeat 3-fold serial dilutions 5 additional times to generate 7 Calibrators.
- 3) Reserve 200 μL of Metabolic Assay Working Solution to be used as zero calibrator.
- 4) Diluted Calibrators should be kept on ice prior to addition to the plate.

Preparation of Serum and Plasma Samples

- 1) The assay format requires 40 μL of sample per well. An adequate volume of each sample should be prepared depending upon desired number of replicates.
- 2) There are numerous proteases in serum and plasma that may cause degradation of GLP-1 and Glucagon. Blood samples should be drawn into tubes containing 500 KIU Aprotinin per mL of whole blood. Alternately, Aprotinin should be added immediately following blood draw. Invert the blood tube several times to mix the sample.
 - a. To obtain serum, tubes containing Aprotinin should be allowed to clot for 30' on a rocker. Spin the tubes for 10 minutes at 1000 x g (4°C) and aliquot serum into separate tubes and store at -80°C until use. Avoid repeated freeze-thaw (> 2) of these aliquots.
 - b. Plasma samples should be obtained in vacutainer or syringe containing Na₂EDTA (1.25 mg/mL) and 500 KIU Aprotinin per mL of whole blood. Tubes should be spun for 10 minutes at 1000 x g (4°C) and then plasma immediately aliquotted into separate tubes and stored at -80°C until use. Avoid repeated freeze-thaw (> 2) of these aliquots.
- 3) Keep isolated or thawed serum/plasma samples on ice or at 4°C prior to subsequent processing or until use in the assay.
- 4) Samples with hemolysis or significant lipemia may hinder accurate assay measurements.

Prepare Detection Antibody Solution

The Detection Antibodies are provided as a 100X stock solution. The working Detection Antibody Solution should contain 1X as final concentration of each antibody.

In a 15 mL tube combine (per plate):

- 90 μL of 10% Blocker D-B
- 30 μL of 100X SULFO-TAG Anti-GLP-1 (7-36)amide Antibody
- 30 μL of 100X SULFO-TAG Anti-m/r Insulin Antibody
- 30 μL of 100X SULFO-TAG Anti-Glucagon Antibody
- 2820 μL of Diluent 100

Prepare Read Buffer

The Read Buffer should be diluted in deionized water to make a final concentration of 1X Read Buffer T. Add 5 mL of stock Read Buffer T (4X) to 15 mL of deionized water for each plate.

Prepare MSD Plate

This plate has been pre-coated with antibodies for the analytes shown in Figure 1. The plate can be used as delivered; no additional preparation (e.g., pre-wetting) is required. The plate has also been exposed to a proprietary stabilizing treatment to ensure the integrity and stability of the immobilized antibodies.

VII Assay Protocol

assay protocol

1. **Addition of Blocker A Solution:** Dispense 150 μ L of Blocker A Solution into each well. Seal the plate with an adhesive plate seal and incubate for 1 hour with vigorous shaking (300–1000 rpm) at room temperature.
2. **Wash and Addition of Sample or Calibrator:** Wash the plate 3X with PBS-T. First, dispense 20 μ L of Metabolic Assay Working Solution into each well of the MSD plate. Then, immediately add 40 μ L of sample or Calibrator into the appropriate wells of the MSD plate. Seal the plate with an adhesive plate seal and incubate for 2 hours with vigorous shaking (300–1000 rpm) at room temperature.
3. **Wash and Addition of the Detection Antibody Solution:** Wash the plate 3X with PBS-T. Dispense 25 μ L of the 1X Detection Antibody Solution into each well of the MSD plate. Seal the plate and incubate for 1 hour with vigorous shaking (300–1000 rpm) at room temperature.
4. **Wash and Read:** Wash the plate 3X with PBS-T. Add 150 μ L of 1X Read Buffer T to each well of the MSD plate. Analyze the plate on the SECTOR Imager. Plates may be read immediately after the addition of Read Buffer.

Notes

Shaking a 96-well MSD MULTI-SPOT plate typically accelerates capture at the working electrode.

Bubbles in the fluid will interfere with reliable reading of MULTI-SPOT plate. Use reverse pipetting techniques to insure bubbles are not created when dispensing the Read Buffer.

VIII Analysis of Results

analysis of results

The calibrators should be run in duplicate to generate a standard curve. The standard curve is modeled using least squares fitting algorithms so that signals from samples with known levels of the analyte of interest can be used to calculate the concentration of analyte in the sample. The assays have a wide dynamic range (3–4 logs) which allows accurate quantitation in many samples without the need for dilution. The MSD Discovery Workbench[®] analysis software utilizes a 4-parameter logistic model (or sigmoidal dose-response) and includes a $1/Y^2$ weighting function. The weighting functionality is important because it provides a better fit of data over a wide dynamic range, particularly at the low end of the standard curve.

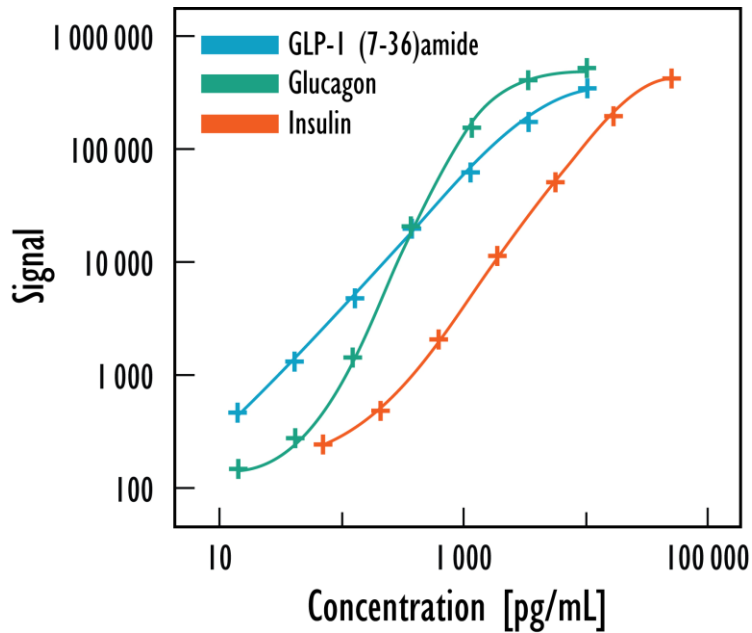
IX

Typical Standard Curve

typical standard curve

The MSD Mouse/Rat GLP-1 (7-36)amide, Insulin, Glucagon Assay is designed for use with mouse or rat serum and plasma samples.

The following standard curves are examples of the dynamic ranges of the assay. The actual signals may vary. A standard curve should be run for each set of samples and on each plate for the best quantitation of unknown samples.



GLP-1 (7-36)amide			Insulin			Glucagon		
Conc. (pg/mL)	Mean	%CV	Conc. (pg/mL)	Mean	%CV	Conc. (pg/mL)	Mean	%CV
0	94	6	0	178	4	0	121	10
14	455	3	69	244	4	14	147	5
41	1287	4	206	473	3	41	277	5
123	4745	2	617	2070	3	123	1399	2
370	19484	5	1852	11625	7	370	20683	5
1111	63575	3	5556	51738	6	1111	154304	6
3333	173685	2	16667	185726	5	3333	400686	3
10000	339390	7	50000	430886	14	10000	498533	3

X Sensitivity

sensitivity

The lower limit of detection (LLOD) is the calculated concentration of the signal that is 2.5 standard deviations over the zero calibrator. Values below represent the average LLODs over multiple kit lots.

	GLP-1 (7-36)amide	Insulin	Glucagon
LLOD (pg/mL)	2.0	39	23

XI Spike Recovery

spike recovery

Serum, heparin plasma, and EDTA plasma were spiked with the calibrators at multiple values throughout the range of the assay. Measured analyte represents average spike recovery in 4-6 pooled mouse serum and plasma. MSD recommends using plasma samples for optimal assay performance. % Recovery = (measured value *100)/expected value

	Average % Recovery		
	GLP-1 (7-36)amide	Insulin	Glucagon
Serum	75	58	74
EDTA Plasma	80	86	99
Heparin Plasma	76	87	96

XII Linearity

linearity

Measured spiked analyte levels in pooled mouse plasma followed by subsequent dilution. % Recovery = (measured value * dilution factor*100)/predicted value

	Average % Recovery		
	GLP-1 (7-36)amide	Insulin	Glucagon
1/2	109	101	113
1/4	113	101	117
1/8	119	98	117

XIII Cross-Reactivity

cross-reactivity

The cross-reactivity shown below is calculated based on signal generated using different GLP-1 isoforms.

GLP-1 (7-36)amide	
Form	Cross-Reactivity
GLP-1 (7-36)amide	100%
GLP-1 (9-36)amide	52%
GLP-1 (1-36)amide	97%
GLP-1 (7-37)	<1%
GLP-1 (1-37)	<1%

XIV Kit Components

kit components

GLP-1 (7-36)amide

Calibrator source: Synthetic amidated peptide (amino acids 7-36) of human GLP-1

Capture Antibody	
Analyte	GLP-1
Source	Mouse monoclonal
Isoforms Recognized	Reacts with all forms of GLP-1, including precursor
Species cross-reactivity	Human, mouse, rat (100% conserved in all mammalian species)

Detection Antibody	
Analyte	GLP-1
Source	Mouse monoclonal
Isoforms Recognized	Reacts with the amidated C terminus of GLP-1 (7-36)amide, GLP-1 (9-36)amide and GLP-1 (1-36)amide, does not react with GLP-1 (7-37), GLP-1 (9-37) or GLP-1 (1-37)
Species cross-reactivity	Human, mouse, rat (100% conserved in all mammalian species)

Insulin

Calibrator source: Recombinant human insulin

The insulin Calibrator has been anchored and referenced to international standards. The table below summarizes the reference information.

Analyte	WHO Standard Reference Number	WHO Standard Units / μg	MSD Calibrator $1\mu\text{g} = \text{WHO Units}$	WHO Units
Insulin	66/304	0.023	0.023	IU

Capture Antibody	
Analyte	Mouse/Rat insulin
Source	Mouse monoclonal
Isoforms Recognized	Does not react with human proinsulin, rat or human C-peptide
Species cross-reactivity	Human, mouse, rat, porcine, bovine

Detection Antibody	
Analyte	Mouse/Rat insulin
Source	Mouse monoclonal
Isoforms Recognized	Does not react with human proinsulin, rat or human C-peptide
Species cross-reactivity	Human, mouse, rat, porcine, bovine

Glucagon

Calibrator source: Synthetic human glucagon (amino acids 1-29)

Capture Antibody	
Analyte	Human glucagon
Source	Mouse monoclonal
Isoforms Recognized	n/a
Species cross-reactivity	Human, mouse, rat

Detection Antibody	
Analyte	Human glucagon
Source	Mouse monoclonal, ascites
Isoforms Recognized	Pancreatic glucagon, reacts weakly to gut glucagon
Species cross-reactivity	Human, mouse, rat, sheep, rabbit, pig, canine, pig, guinea pig

GLP-1

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Summary Protocol
MSD 96-well MULTI-ARRAY
Mouse/Rat GLP-1 (7-36)amide, Insulin, Glucagon Assay Kit

MSD provides this summary protocol for your convenience.
Please read the entire detailed protocol prior to performing
the Mouse/Rat GLP-1 (7-36)amide, Insulin, Glucagon Assay.

Step 1 : Sample and Reagent Preparation

Bring appropriate Diluents and plates to room temperatures.

Prepare Blocker A Solution.

Prepare serum or plasma samples.

Prepare Metabolic Assay Working Solution and keep on ice.

The Calibrator stock solutions should be thawed and kept on ice.

Prepare an 8-point standard curve using supplied calibrator:

- The Calibrator should be diluted in Metabolic Assay Working Solution.
- Dilute the stock Calibrators 1:100 as indicated in Reagent Preparation section, then perform a series of 3-fold dilution steps and a no calibrator blank.
- Diluted Calibrators should be kept on ice until use.

Prepare Detection Antibody Solution by diluting the 100X Detection Antibodies into a1X final concentration of each antibody. The Detection antibodies should be diluted in 3.0 mL of Diluent 100 containing Blocker D-B as indicated in Reagent Preparation section.

Prepare 20 mL of 1X Read Buffer T by diluting 4X MSD Read Buffer T (with surfactant) with deionized water.

Step 2 : Add Blocker A Solution

Dispense 150 μ L/well Blocker A Solution.

Incubate at room temperature with vigorous shaking (300–1000 rpm) for 1 hour.

Step 3 : Wash and Add Sample or Calibrator

Wash plate 3X with PBS-T.

Dispense 20 μ L/well Metabolic Assay Working Solution.

Immediately, dispense 40 μ L/well Calibrator or Sample.

Incubate at room temperature with vigorous shaking (300–1000 rpm) for 2 hours.

Step 4 : Wash and Add Detection Antibody Solution

Wash plate 3X with PBS-T.

Dispense 25 μ L/well 1X Detection Antibody Solution.

Incubate at room temperature with vigorous shaking (300–1000 rpm) for 1 hour.

Step 5 : Wash and Read Plate

Wash plate 3X with PBS-T.

Dispense 150 μ L/well 1X Read Buffer T.

Analyze plate on SECTOR instrument.

